FIVE NEW LABDANE DITERPENE OXIDES FROM EUPATORIUM JHANII*

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Abstract—Five new labdane diterpene oxides jhanol, jhanidiol, jhanol acetate, jhanidiol-18-monoacetate and jhanidiol diacetate were isolated from the aerial parts of *Eupatorium jhanii*.

INTRODUCTION

From species of *Eupatorium*, several sesquiterpene lactones with cytotoxic activity [1-67] and a number of interesting flavonoid compounds [7-11] have been isolated. However, there is not much information about the presence of diterpenoid compounds in this genus [12,13].

In the present communication, we report the isolation and structural determination of five hydroxylated derivatives of 8,13-epoxylabd-14-ene (manoyl oxide) from Eupatorium jhanii.

RESULTS

The first free alcohol isolated was jhanol, $C_{20}H_{34}O_2$, (1a). Spectral and chemical data showed it to be a primary alcohol: IR 3480 cm⁻¹, ¹H NMR two doublets, J 11 Hz at 3.14 and 3.45; it gave an acetate and was oxidized to the aldehyde 1c.

The presence of a vinyl group, suggested by the IR spectrum (bands at 3080, 1640, 980 cm⁻¹), was confirmed by the ¹H NMR spectrum which exhibited the signals characteristic of this group: a set of three quartets at 4.95, 5.15 and 5.92 ppm [14].

The remaining oxygen atom must be part of an ether bridge connecting two fully substituted carbon atoms; the IR spectrum showed strong absorption at 1120 cm⁻¹ while there was no ¹H NMR signal for a hydrogen geminal to the oxygen. This spectrum also showed two methyl singlets at 1.31 and 1.35 ppm (Me—C(0)) and two more at 0.80 and 0.88 ppm. Consequently jhanol is probably a compound belonging to the 8,13-epoxylabdene series.

This was proved by Huang-Minlon reduction of the aldehyde 1c, affording two substances identified as 8,13-epoxylabdane (2a) [15,16] and 8,13-epoxylabd-14-ene

Table 1. ¹³C-NMR spectra of manoyl oxide (1d) jhanol acetate (1b) and jhanidiol-18-monoacetate (3b)

Carbon No	1d	1 b	3ь
1	39.12	38.61	79.14
2	18.62	17.20	28.89
3	42.23	35.93	34.04
4	33.32	36.56	36.22
5	56.55	50.64	49.26
6	20.03	19.95	19.73
7	43.40	43.01	42,77
8	75.09	74.89	74.80
9	55.60	56.05	55.86
10	37.11	37.02	42,91
11	51.41	15.42	18.49
12	35.73	35.93	35.30
13	73.25	73.28	73.28
14	148.07	148.04	147.96
15	110.21	110.23	110.28
16	28.71	78.52	28.81
17	25.58	25.43	25.69
18	33.40	72.99	72.49
19	21.39	17.80	16.91
20	15.41	15.83	11.71

^{*} Part 33 in the series "Chemistry of the Compositae". For Part 32 see: A. G. González, J. Bermejo, J. L. Bretón, G. Martínez, B. Domínguez, J. M. Amaro, J. Chem. Soc. Perkin I 1976, 1663.

⁽¹d) [17,18]. Comparison of the methyl carbon-13 NMR signals of the jhanol acetate with those of manoyl oxide (2a), Table 1, showed that 1b is an 18-hydroxyditerpene. The C-18, C-19 and C-20 methyls of manoyl oxide [19,20] and also of other tri and tetracyclic diterpenes resonate at different field positions, near to 33, 20 and 15 ppm [21-23]. On the other hand, the ¹³C-NMF (SFORD) spectrum of 1b showed only two quartets, at 17.80 and 15.23 ppm, and a new triplet at 72.5 ppm. The stereochemistry at C-4, depicted in 1b, was corroborated by the ¹H-NMR spectrum because both the methyl and oxymethylene signals occupy different field positions in the two possible structures [24,25]. The position of the aldehyde signal for 1c is typical of equatorial substituents [26].

The most polar diterpene of this series, jhanidiol (3a), had the empirical formula $C_{20}H_{34}O_3$; its polarity and the ¹H-NMR spectrum, which showed signals for protons under the hydroxyls at 3.14d, 3.45d and 3.41t ppm, suggested the presence of a primary and a secondary alcohol; according to the conditions, this compound gave the monoacetate 3b or the diacetate 3c. Comparison of the ¹H-NMR resonances of 1a and 3a made it obvious

¹H-NMR signals for a Δ^2 -1-one [29], in full accord with structure 3a. Jhanol acetate (1b), jhanidiol-18-monoacetate (3b) and jhanidiol diacetate (3c) were also found as natural products in the plant.

EXPERIMENTAL

Mp's are uncorr. Optical activities were taken in CHCl₃, ¹H-NMR spectra on a 90 MHz and ¹³C-NMR on a 20 MHz

that the only difference between these compounds was the existence of an additional OH group in 3a. Oxidation of monoacetate 3b to the ketone 3d and Huang-Minlon reduction yielded an alcohol identical with jhanol (1a).

Analyses of the ¹³C-NMR spectrum of 3b indicated the position and stereochemistry of oxymethine hydrogens. In the expoylabdane [19,20] and also in other series of diterpenes, C-2, C-6 and C-11 triplets were found near 20 ppm; the resonance signals of C-1, C-3, C-7 and C-12 appeared much lower in the field [21-23]. Comparison of the secondary carbon resonances of 1b and 3b suggested that one of the latter's carbons must be substituted The paramagnetic shift undergone by C-2 (11.7) and C-10 (5.9), together with the diamagnetic one experienced by C-3 (11.9) and C-20 (4.12 ppm) clearly point to C-1 as the locus of the hydroxyl group with an equatorial orientation [27]. The ¹H-NMR spectrum of 3c corroborated this conclusion, showing the oxymethine resonance as a clear quartet at 4.53 ppm. Moreover, the spectra of all the oxidation products of 3a had signals for the equatorial proton at C-11 [28].

Jhanidiol did not form an acetonide nor did the ketoacid 3f show β -ketoacid properties. Bromination and dehydrobromination of ketodihydroacetate 2e afforded the enone 4 which presented the characteristic

instrument in CDCl₃ with TMS as internal reference. Column and dry column chromatography was performed on Si gel.

Isolation of the diterpenes. Air-dried aerial part of the plant Eupatorium jhanii Robinson (4 kg) collected at El Delgadito (Mérida, Venezuela) in June, was finely cut and Soxhlet extracted several times with EtOH. Combined filtered ethanolic extracts were concentrated in vacuo and chromatographed on a Si gel column. Petrol, petrol-EtOAc and EtOAc eluted a mixture of diterpenes, triterpenes and flavones which was rechromatographed on dry column, yielding the following diterpenes, in the order of elution: jhanol acetate (1b), jhanidiol diacetate (3c), jhanidiol-18-monoacetate (3b), jhanol (1a), jhanidiol (3a).

Jhanol acetate (1b). 1b (400 mg) was isolated as a liquid; IR $v_{\rm max}^{\rm CHCl_3}$ cm⁻¹: 3080, 1720, 1640, 1120, 990, 920; NMR δ 0.88 (6H, s, C₁₀C₂₀), 1.31 and 1.35 (each 3H, s, C-14 and C-17), 2.10 (3H, s), 3.68, 3.90 (each 1H, d, J 11 Hz, C-18). 4.97 (1H, q, J 11 Hz, J 2 Hz), 5.17 (1H, q, J 17 Hz, J 2 Hz), 5.94 (1H, q, J 11 Hz, J 17 Hz); MS: m/e (%) 348 (1) (M⁺), 333, 273, 255, 191, 149 (100), 135. Identical to that obtained from jhanol by standard acetylation.

Jhanidiol acetate (3c). 3c (500 mg was isolated as a liquid; IR $\nu_{\rm max}^{\rm CHCl_3}$ cm⁻¹: 3080, 1720, 1110, 980, 920; NMR. δ 0.80 (3H, s), 0.96 (3H, s), 1.20 (3H, s), 1.26 (3H, s), 1.96 (3H, s), 2.02 (3H, s), 3.67, 3.84 (each 1H, d, J 11 Hz), 4.53 (1H, q, J 9 and 5 Hz), 4.94 (1H, q, J 11 and 2 Hz), 5.12 (1H, q, J 17 and 2 Hz), 5.86 (1H, q, J 17 and 11 Hz). Identical to the product obtained by acetylation of jhamdiol. MS: m/e (%) 391 (M⁺-15), 331, 271, 253, 223, 201, 189, 176, 174, 149 (100), 133.

Jhanidiol-18-monoacetate (3b). 3b (1200 mg) was isolated as a liquid; IR $v_{\rm max}^{\rm KBr}$ cm $^{-1}$: 3330, 3080, 1740, 1640, 1240, 1120, 980, 920; NMR: δ 0.82, 0.88, 1.24, 1.29, 2.06 (all 3H, s), 3.38 (1H, q, $W_{1/2}$ 14 Hz), 3.62, 3.84 (each 1H, d, J 11 Hz), 4.92 (1H, q, J 11 and 2 Hz), 5.14 (1H, q, J 17 and 2 Hz), 5.80 (1H, q, J 17 and 11 Hz); MS: m/e ($\frac{9}{2}$) 364 (1) (M $^+$), 349 (100), 331, 313, 271, 253, 223, 201, 188, 149.

Jhanol (1a). 1a (2.3 g) was isolated as a solid; mp 139–141° $[\alpha]_D + 27^\circ$ (c 0.65); IR $\nu_{\text{max}}^{\text{KBr}}$ cm⁻¹: 3480, 3080, 1640, 1120, 980, 930; NMR · δ 0.80, 0.88, 1.31, 1.35 (each 3H, s), 3.15, 3.47 (each 1H, d, J 11 Hz), 4.95 (1H, q, J 11 and 2 Hz), 5.15 (1H, q, J 17 and 2 Hz), 5.92 (1H, q, J 17 and 11 Hz); MS: m/e (%) 306 (1) (M⁺), 291, (98), 273, 261, 255, 243, 208, 177 (100), 151, 149, 147, 135, 133, 123, 121. (Found: C78.30; H 10.89.C₂₀H₃₄O₂ requires: C 78.38; H 11.18%).

Jhanidiol (3a). 3a (4.0 g) was isolated as a solid; mp 189° [α]_D +90° (c 0.8); IR $\nu_{\rm max}^{\rm KBr}$ cm⁻¹ 3460, 3080, 1640, 1120, 990, 920; NMR: δ 0.78, 0.92, 1.28, 1.34 (each 3H, s), 3.14, 3 45 (each 1H, d, J 11 Hz), 3.41 (1H, t, W_{1/2} 14 Hz), 4.95 (1H, q, J 11 and 2 Hz), 5.15 (1H, q, J 17 and 2 Hz); MS: m/e (%) 322 (0.5), (M⁺), 307 (100), 289, 271, 259, 231, 224, 206, 193, 189, 175, 173, 161, 159, 149, 137. (Found: C 74.18; H 10.71. C₂₀H₃₄O₃ requires: C 74.49: H 10.63%).

Oxidation of jhanol. Pyridinium chlorochromate (1.5 g) was suspended in dry CH₂Cl₂ (20 ml) and jhanol (290 mg) in CH₂Cl₂ (15 ml) was added once to the stirred sol. After 1 hr, dry Et₂O (20 ml) was added and the supernatant decanted from the black gum formed. The insoluble residue was washed $3 \times$ with 30 ml portions dry Et₂O. Combined CH₂Cl₂-ether sol was passed through a short column of Celite and solvent removed by distillation, giving 8,13-epoxylabd-14-en-al (210 mg) (1c); IR $v_{\text{max}}^{\text{CHCl}_3}$ cm⁻¹: 3080, 2700, 1720; NMR: δ 0.85, 1.04, 1.28, 1.31 (each 3H, s), 4.94 (1H, q, J 11 and 2 Hz), 5.12 (1H, q, J 17 and 2 Hz), 5.89 (1 H, q, J 17 and 11 Hz) and 9.18 (1 H, s).

of 8,13-epoxylabdan-18-al. reduction Huang–Minlon N₂H₄.H₂O was added to a soln of 1c (210 mg) in diethylene glycol (14 ml) and the mixture refluxed for 90 min (temp. 130°). After the addition of KOH pellets (300 mg), refluxing continued for a further 45 min. Then the temp was raised to 210° by distillation of the excess water-hydrazine and the refluxing continued for another 3 hr. The usual work-up afforded a mixture of 2 products which were separated by dry column chromatography, using petrol-C₆H₆ (4:1) as eluent. 8,13-Epoxylabdane (dihydromanoyl oxide) (2a) (65 mg); isolated as a liquid $[\alpha]_D + 3^\circ (c \ 1.1)$ Lit + 5.9; NMR: δ 0.78, 0.80, 0.85 (t), 0.86, 1.18, 1.29 (each 3H, s) 8, 13-Epoxylabd-14-ene (manoyl oxide) (1d). 1d (95 mg) was isolated as a solid; mp $26-28^{\circ}$ [α]_D + 11 (c 2.0) Lit 22.5-26.5 $[\alpha]_D + 19$; NMR; δ 0.83 (6H, a), 0.90, 1.31, 1.33 (each 3H, s), 4.94 (1H, q, J 11 and 2 Hz), 5.16 (1H, q, J 17 and 2 Hz), 5.92 (1H, q, J 17 and 11 Hz); MS: m/e (%) 290 (1) (M+), 275 (100), 257, 245, 192, 177, 149, 137.

Partial acetylation of jhanidiol. To a soln of 3a (700 mg) in C_5H_5N (2 ml), cold Ac_2O (3 ml) was added and the mixture was left for 10 min. The usual work-up gave a mixture of diacetate (70 mg), monoacetate (410 mg) and unaltered starting material, which was separated by dry column chromatography, using C_6H_6 -EtOAc (9:1) as eluent. The diacetate and 18-monoacetate of jhanidiol were identical to the natural products obtained.

Oxidation of jhanidiol monoacetate (3b). 3b (410 mg) was oxidized as described above for 1a. 18-acetate-1-keto-8,13-epoxylabd-14-ene (400 mg) was obtained; mp 87-89°; $[\mathbb{Z}]_{D}$ +70°; $[\mathbb{R} \ \nu_{\max}^{KBr} \ cm^{-1}$: 3090, 1735, 1700, 1240, 1120, 990, 920, 910; $[\mathbb{N}MR: \delta \ 1.05, 1.21, 1.30, 1.35$ (each 3H, s), 2.06 (3H, s), 2.8 (1H, m), 3.74, 3.98 (each 1H, d), 4.93 (1H, q), 5.13 (1H, q), 5.93 (1H, q); $[\mathbb{N}MR: \delta \ (\text{benzene-d}_{\delta})\ 0.72, 0.92, 1.20, 1.36$ (each 3H, s), 3.58, 3.76 (each 1H, d), 4.93 (1H, q), 5.26 (1H, q), 5.97 (1H, q); $[\mathbb{N}S: m/e\ (\%)\ 362\ (1)\ (\mathbb{M}^+), 347, 269, 224, 200, 191, 173, 167, (100).$

Jhanol from jhanidiol. The above substance 3d (300 mg) was reduced under Huang-Minlon conditions. Work up gave jhanol: mp, mixed mp and IR superimposable.

Hydrogenation of 3d. 3d (400 mg) was dissolved in EtOAc

(20 ml) and hydrogenated for 1 hr over Pd/C(5 %) at room temp and atm press. This gave 2c (350 mg); NMR: δ 0.85 (3H, t, J 6 Hz), 1.04 (3H, s), 1.21 (6H, s) and 1.32 (3H, s).

Bromination of keto-18-acetoxy-8,13-epoxy-labdane. A soln of trimethylphenylammonium tribromide (400 mg) in dry THF (6.ml) was added in drops while stirring a soln of 2e (350 mg) in the same solvent until the mixture turned yellow and it was then stirred at room temp for a further 30 min. The reaction mixture was poured into $\rm H_2O$, extracted with $\rm CH_2Cl_2$ and the solvent was evaporated, yielding a Br derivative (300 mg) which did not crystallize; IR $\nu_{\rm max}^{\rm CHCl_3}$ cm⁻¹: 1730, 1700, 1120, 960; NMR: δ 0.82 (3H, t, 6 Hz), 1.02 (3H, s), 1.18 (6H, s), 1.30 (3H, s), 2.03 (3H, s), 2.8 (1H, m), 3.64, 3.72 (each 1H, d), 4.12 (1H, m $W_{1/2}$ 27 Hz).

Dehydrobromination of 1-keto-2β-bromo-18-acetoxy-18,13-epoxylabdane. A mixture of Li₂CO₃ (400 mg), LiBr (250 mg), DMF (15 ml) and C₆H₆ (6 ml) was heated until the H₂O and C₆H₆ had been eliminated. Then a soln of bromide (300 mg) in DMF (15 ml) was added and the reaction mixture refluxed for 1 hr under N₂. The cooled soln was poured into H₂O, extracted with CH₂Cl₂ and washed with 10% aq HOAc, sat aq NaHCO₃ and H₂O. After the solvent was evaporated, the residue afforded 1-keto-18-acetoxy-8,13-epoxy-labd-2-ene (4); mp 84-88°; $[\alpha]_D$ +46°; UV $\lambda_{\max}^{\text{EICH}}$ mm: 223 (c 22000); IR $\nu_{\max}^{\text{CHCl}_3}$ cm⁻¹; 1730, 1700, 1680, 1120, 950; NMR; δ 0.84 (3H, t J6 Hz), 1.02 (3H, s), 120 (6H, s), 1.32 (3H, s), 2 03 (3H, s), 2.75 (1H, m, W_{1/2} 27 Hz), 3.71, 3.50 (each 1H, d, J 11 Hz), 5.82 and 6.32 (each 1H, d, J 11 Hz),

8,13-Epoxy-labd-14-en-1-keto-18-al (3e). 3a (100 mg) was oxidized as described above for 1a, giving 3e (80 mg); mp 88-93°; $[\alpha]_D$ +62; M⁺ 318; IR $v_{\rm max}^{\rm BB}$ cm⁻¹. 3080, 2700, 1720, 1700, 1640, 1260, 1130, 990, 960, 920, 895; NMR: δ 1.23, 1.27, 1.30, 1.35 (each 3H, s), 2.78 (1H, m), 4.92 (1H, q), 5.11 (1H, q), 5.86 (1H, q).

Oxidation of keto-aldehyde (3f). 3e (100 mg) dissolved in Me_2CO (min quantity) was treated dropwise with Jones' reagent and left at room temp for 3 hr, upon which MeOH was added to destroy any excess reagent. The mixture was poured into water and extracted as usual, giving 3f; mp $206-209^\circ$; $[\alpha]_D + 33^\circ$ (c 0.57); $IR \ \nu_{max}^{CHCl_3}$ cm $^{-1}$: 3500–2500 (br) 1700, 1640, 1110; NMR: δ 1.17, 1.29 (each 3H, s), 4.90 (1H, q), 5.12 (1H, q), 5.88 (1H, q); MS: m/e (%) 319 (100) (M $^+$ -15), 256, 186, 173, 149.

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REFERENCES

- Kupchan, S. M., Kelsey, J. E., Maruyama, M., Cassidy, J. M., Hemingway, S. C. and Knox, J. R. (1969) J. Org. Chem. 34, 3876.
- Kupchan, S. M., Fujita, T., Maruyama, M. and Britton, R. W. (1973) J. Org. Chem. 38, 1260.
- 3. Herz, W. and Wahlberg, I. (1973) J. Org. Chem. 38, 2485.
- Kupchan, S. M., Maruyama, M., Hemingway, R. J. Shibuya, S. and Fujita, T. (1973) J. Org. Chem. 38, 2189.
- 5. McPhail, A. T., Onan, R. D., Lee, K. H., Ibuka, T. and Huang, H. C. (1974) Tetrahedron Letters 3203.
- 6. Herz, W. and Sharma, R. P. (1976) Tetrahedron 41, 1015.
- Kupchan, S. M., Sigel, W., Knox, J. R. and Udayamurthy, M. S. (1969) J. Org. Chem. 34, 1460.
- 8. Wagner, H. Iyengar, M. A., Hörhammer, L. and Herz, W. (1972) Phytochemistry 11, 1504.
- Wagner, H., Iyengar, M. A., Seligmann, O., Hörhammer, L. and Herz, W. (1972) Phytochemistry 11, 1506.
- Herz, W., Gibaja, S., Bhat, S. V. and Srinivasan, A. (1972) Phytochemistry 11, 2859.
- Talapatra, S. R., Bhar, D. S. and Talapatra, B. (1974) Australian J. Chem. 27, 1137.

- Hegnauer, R. (1964) Chemotaxonomie der Pflanzen, Vol. III. p. 482. Birkhauser, Basel.
- Herz, W. and Sharma, R. P. (1976) J. Org. Chem. 41, 1021.
- 14. Wenkert, E., Beak, P. and Grant, P. K. (1961) Chem. Ind (London) 1574.
- Hosking, J. R. and Brandt, C. W. (1934) Ber Deut. Chem. Ges 67, 1173.
- Hasen, A. J., Kimland, B., Almqvist, S. O. and Enzell, C. R. (1972) Acta Chem. Scand. B26, 832.
- 17. Hodges, R. and Reed, R. I. (1960) Tetrahedron 10, 71.
- 18. Giles, J. A., Schumacher, J. N., Mims, S. S. and Bernasek, E. (1962) Tetrahedron 18, 169.
- Buckwalter, B. L., Burfitt, I. R., Nagel, A. A. and Wenkert, E. (1975) Helv. Chim. Acta 58, 1567.
- Almqvist, S. O., Enzell, C. R. and Wehrli, F. N. (1975) Acta Chem. Scand. B29, 695.
- 21. Carstenn-Lichterfelde, C., Pascual, C., Pons, J., Rabanal, R.,

- Rodríguez, B. and Valverde, S. (1975) Tetrahedron Letters 3569
- 22. Hanson, J. R., Siverns, M., Piozzi, F. and Savona, G. (1976) J. Chem. Soc. Perkin I 114.
- Wahlberg, I., Almqvist, S. O., Nishida, I. and Enzell, C. R. (1975) Acta Chem. Scand. B29, 1047.
- Wenkert, E., Afonso, A., Beak, P., Carney, R. V. J., Jeffs,
 P. N. and McChesney, J. D. (1965) J. Org. Chem. 30, 713.
- Gaudemar, A., Polonsky, J. and Wenkert, E. (1964) Bull. Soc. Chim. France 407.
- 26. King, T. J. and Yarley, J. P. (1961) J. Chem. Soc. 4308.
- Eggert, H., Van Antwerp, G. F., Bhacca, N. S. and Djerassi,
 C. (1976) J. Org. Chem. 41, 71.
- Bhacca, N. S. and Williams, D. H. (1966) Application of NMR Spectroscopy in Organic Chemistry, p. 66. Holden Day, San Francisco.
- Chan, W. R., Gibbs, J. A. and Taylor, D. R. (1973) J. Chem. Soc. Perkin I 1047.